





# **Standard Treatment Workflow (STW) of**

## MICROBIOLOGICAL WORK-UP FOR ADULT EXTRAPULMONARY TUBERCULOSIS

#### LOGISTICS INVOLVED IN SAMPLE COLLECTION AND TRANSPORTATION

- Collect Samples for Microbiological work-up in sterile containers before treatment is started. (Mention date & time of collection)
- Specimens to be sent in sterile saline (NOT in formalin)
- Establish linkages between peripheral centres, District centres and Tertiary centre/medical colleges/ IRL. Specify details of person to be contacted, department and contact number during referrals
- Transportation at 2-8 °C
- Maximum time for transportation in cold chain should be 5 days from time of collection
- Quantity of sample mentioned is only for microbiological work-up. Tests like histopathology, cytology, ADA, glucose, protein, etc will require additional sample
- Microbiological tests for TB (smear, molecular tests, culture) will be performed as per availability and preparedness of site
- PHC and CHC should perform smear microscopy and molecular diagnostic tests. If sample less than 500 µl, refer directly to Tertiary centre/medical colleges/IRL for culture. Residual sample in the needle and syringe used to collect the specimen can be used for smear
- MGIT to be used for culture. However, if MGIT is not available, LJ medium should be used

#### **REJECTION OF SAMPLES**

- Unlabelled samples (All specimens) MUST be labelled & have a unique patient identifier)
- Have no collection date indicated
- Insufficient quantity No specimen in container
- Damaged Specimen leaked or broken in transit
- Samples greater than 3 days old at room temperature and more than 5 days in refrigeration are unreliable specimens for testing

Precious samples should be transported to IRL.

**Diagnostic algorithm of NTEP to be** followed in the Microbiology labs

### **MICROBIOLOGICAL GUIDANCE FOR COMMON TYPES OF EXTRAPULMONARY TUBERCULOSIS**

OSTEOARTICULAR/ MUSCULOSKELETAL	<ul> <li>Sample: Tissue, pus, synovial fluid</li> <li>Sample amount: Biopsy: Specimen material 1 cm x 1 cm biopsies. Any caseous area should be sampled. Add 0.5-2 ml sterile saline to biopsy depending on its size to avoid drying of tissue specimen</li> <li>Optimum fluid/pus: 2-3ml.</li> <li>Swabs are sub-optimal samples</li> </ul>	Pr • Pr nc at • If s	<ul> <li>Processing:</li> <li>Preferably immediately. If not possible- store/transport at 2-8 °C</li> <li>If sample is adequate, attempt molecular testing at that site</li> <li>If biopsy is not possible or at an inaccessible site, refer patient to the next higher centre immediately where appropriate test can be done</li> </ul>
PLEURAL	• Sample: Pleural fluid • Sample amount: 10-15 ml		
INGITIS	• Sample: CSF:		

S MENIN	• Sample amount: 3-5 ml	<ul> <li>If sample obtained at a centre is inadequate, send directly to nearest Tertiary centre/medical colleges/IRL</li> </ul>
LYMPHADENIT	<ul> <li>Sample: FNA/ Biopsy</li> <li>Sample amount: Specimen material 1 cm x 1 cm biopsy. Add 0.5-2 ml sterile saline to biopsy depending on its size to avoid drying of tissue specimen</li> <li>Optimum FNA sample: 2 ml</li> </ul>	
UROGENITAL	<ul> <li>Sample: urine</li> <li>Sample amount: Entire early morning urine sample (3-5 days)</li> </ul>	<ul> <li>Microbiological procedures:</li> <li>AFB Smear Microscopy except in GI TB</li> <li>NAAT</li> <li>Culture (MGIT. If MGIT is not available LJ medium should be used)</li> <li>Drug susceptibility testing, if culture is positive</li> </ul>
FEMALE GENITAL	<ul> <li>Sample: Endometrial curettage/biopsy</li> <li>Sample amount: Biopsy: Specimen material 1cm x 1 cm biopsies. Any caseous area should be sampled. Add 0.5-2 ml sterile saline to biopsy depending on its size to avoid drying of tissue specimen</li> </ul>	
GASTROINTESTINAL	<ul> <li>Sample: Tissue, pus, peritoneal fluid</li> <li>Sample amount: Biopsy: Specimen material 1 cm X 1 cm biopsy (Atleast 6 biopsies for microbiological diagnosis including any caseous area). Any caseous area should be sampled. Add 0.5-2 ml sterile saline to biopsy depending on its size to avoid drying of tissue specimen</li> <li>Optimum fluid/pus: 5-10ml</li> </ul>	

#### **ABBREVIATIONS**

**ADA: Adenosine Deaminase** AFB: Acid fast bacilli **CHC: Community Health Centre** 

FNA: Fine needle aspirate PHC: Primary health Centre LJ medium: Lowenstein Jensen medium **TB**: **Tuberculosis IRL: Intermediate Reference** MGIT: Mycobacteria Growth Indicator tube (Liquid culture medium for mycobacteria) NAAT: Nucleic Acid Amplification Tests-Xpert MTB/RIF/TrueNat laboratory

## **REFERENCES**

1. National TB Elimination Programme, Central TB Division. Training Modules for programme managers & Medical Officers. Ministry of Health & Family Welfare, Government of India. https://tbcindia.gov.in/index1.php?lang=1&level=1&sublinkid=5465&lid=3540 Last access on 15 March. 2022.

2. Guidelines for programmatic management of drug resistant tuberculosis in India March 2021. National TB Elimination Programme, Central TB Division, Ministry of Health & Family Welfare, Government of India accessed at https://tbcindia.gov.in/showfile.php?lid=3590 Last access on 15 March, 2022.

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